

The UK's commitment to the OSPAR Hazardous Substances strategy requires that information is available on the toxicity of hazardous substances present in the marine environment, particularly in marine sediments. In this study, toxicity identification evaluation (TIE) techniques were used to characterise the toxic substances present in sediments collected from six UK estuaries that have been historically impacted by primary industry; Tyne, Tees, Mersey, Humber, Thames and Southampton Water.

Spatial survey

As a first step sediment pore water extracts and sediment solvent extracts were tested using a battery of bioassays. Pore water samples were tested for the presence of lethal toxicants by using the *Tubifex* benthic bioassay, a *Caenorhabditis* receptor (ER) agonists using the yeast *Saccharomyces cerevisiae* (YES) and for *in vitro* mutagenic activity using the *Plasmid*[®] assay (Figure 1; Table 1). Sediment solvent extracts were also tested for ER agonists and mutagens, while *in vitro* hydrocarbon receptor (AhR) agonists were also tested for by using the ER-CAH2000 assay (Figure 1; Table 1). Lethal toxicity was only measured in 100% of the sediment pore water samples tested. ER agonists were detected in 66% of sediment pore water samples and 75% of sediment solvent extracts tested. Pore water samples tested for any of the pore water samples while 25% of sediment solvent extracts scored positive. All of the samples tested using the ER-CAH2000 assay scored positive.



Figure 2. Fractionation scheme for the characterization of marine sediments.

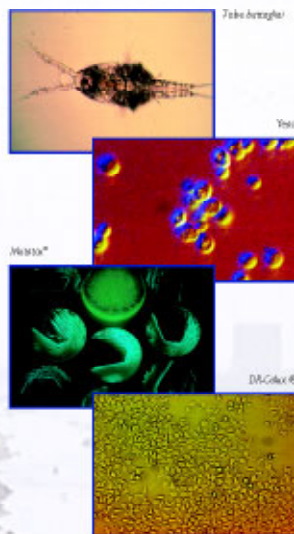


Figure 1: Elements used in the study

Toxicity identification evaluation (TIE)

TE techniques using the above beams to direct the fractionation of the complex environmental samples, were then applied in an attempt to identify the compounds responsible for the observed effects (Figure 2). Amonium ion was responsible for much of the lethal toxicity to *T. testigolus*, although organic metals, unknown hydrocarbons and unknown substances also contributed to the toxicity of one sample. TE techniques were successfully applied to identify polycyclic aromatic hydrocarbons (PAH), polycyclic aromatic isosteres (PAKS), cox-PAH and nitro-PAH as potentially contributing to the mutagenic signal measured in certain sediments. Unknown polar and non-polar mutagens were also shown to be present. Nonylphenol, octrinarine and diethyl-4,4'-di-n-but-2-ene were shown to contribute to the *in vitro* ER agent potency of sediment solvent extracts; however, the identity of the compounds responsible for the majority of the measured ER agent activity in both sediment solvent extracts and pore waters remain unknown. The results of the DR-CAUSE® bioassay showed that diethyl-*p*-dichloro, dibutylaroms and PAH were acting as ER agents in certain samples (Table 2).

Comparison of the identified substances with OSPAR lists of substances for priority action and possible concern showed a number not to be included. Where available, occurrence, persistence, bioaccumulation and toxicity data indicated that certain of these compounds should be considered for further assessment.

Table 2: Summary of compounds identified

Substance	CAS No.	OSN1 listing category			Function or use category	Persistent (y)	Bioaccumulative (log K _{ow})	Toxic	Source
		Priority action	Proactive concerns	No listing					
Nonylphenol	904-60-5	✓	✓	✓	Phenol				Surfactant parabens
Chloroform	298-07-7			✓	Dye	?	?	(<i>in vitro</i> ER potency) ^a (<i>in vitro</i> ER potency) ^a	Dye
Chlorate 4,6-dim-3-one	958-03-0			✓	Chloral and oxidant product	?	?		Household
Biphenyl	90-52-4	✓							
Polystyrene									
Polystyrene	81-20-3	✓			PMU				
1-Methylpiperazine	30-12-0	✓			PMU				
1,2-Dimethylpiperazine	877-09-9	✓			PMU				
1,3-Dimethylpiperazine	898-01-5	✓			PMU				
1,4-Dimethylpiperazine	571-55-4	✓			PMU				
1,2-Dimethylpiperazine	573-05-0	✓			PMU				
1,3,5-Triethylpiperazine	3021-69-1	✓			PMU				
Acetophenone	298-06-9	✓	✓		PMU				
Fluorene	86-73-7	✓			PMU				
9,9-Dimethyl-9H-fluorene	459-45-3	✓			PMU				
9-Methyl-9H-fluorene	86-07-6	✓	✓		PMU				
Phenylacetylene	832-77-9	✓			PMU				
3-Methylphenylacetylene	1570-69-8	✓			PMU				
2,7-Dimethylphenylacetylene	830-64-4	✓			PMU				
1-Methylphenylacetylene	813-12-1	✓			PMU				
Linear tri-n-octyl complex mixture (LCO)		✓			PMU				
Fluoranthene	208-94-0	✓	✓		PMU				
Pyrene	129-00-0	✓	✓		PMU				
1-Methylpyrene	2301-21-7	✓			PMU				
4-Methylpyrene	3355-12-6	✓			PMU				
1,2-Dimethylpyrene	6481-21-4	✓			PMU				
1,9-Dimethylpyrene	249-17-4	✓			PMU				
Benzofluorene	55-55-3	✓			PMU				
4-Methylbenzofluorene	2301-31-8	✓			PMU				
5-Methylbenzofluorene	229-95-2	✓			PMU				
1-Methylfluorene	3001-39-8	✓			PMU				
2-Methylfluorene	4332-71-3	✓			PMU				
2,3-Dimethylfluorene	812-76-2	✓			PMU				
Fluoranthene	208-94-0	✓	✓		PMU				
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1,9-Dimethylpyrene									

^aAs included under the general heading of polyacyl derivative hydrolases, EC 3.1.1.1, also catalyzes polyacyl 3.3.1.11¹ when compared to EC 3.1.1.1² and polyacyl 3.1.1.1³ when compared to EC 3.1.1.1⁴ and EC 3.1.1.1⁵ (data not shown). ¹RISS, ²RISS, ³RISS, ⁴RISS, ⁵RISS. ⁶Found in 59 included polyacyl hydrolase in this study. ⁷Marigold was used as the substrate in the Marigold assay. ⁸W60, ⁹Salmon-Salmon, ¹⁰W60, ¹¹Salmon-Salmon, ¹²W60, ¹³Salmon-Salmon, ¹⁴W60, ¹⁵Salmon-Salmon, ¹⁶W60, ¹⁷Salmon-Salmon, ¹⁸W60, ¹⁹Salmon-Salmon, ²⁰W60, ²¹Salmon-Salmon, ²²W60, ²³Salmon-Salmon, ²⁴W60, ²⁵Salmon-Salmon, ²⁶W60, ²⁷Salmon-Salmon, ²⁸W60, ²⁹Salmon-Salmon, ³⁰W60, ³¹Salmon-Salmon, ³²W60, ³³Salmon-Salmon, ³⁴W60, ³⁵Salmon-Salmon, ³⁶W60, ³⁷Salmon-Salmon, ³⁸W60, ³⁹Salmon-Salmon, ⁴⁰W60, ⁴¹Salmon-Salmon, ⁴²W60, ⁴³Salmon-Salmon, ⁴⁴W60, ⁴⁵Salmon-Salmon, ⁴⁶W60, ⁴⁷Salmon-Salmon, ⁴⁸W60, ⁴⁹Salmon-Salmon, ⁵⁰W60, ⁵¹Salmon-Salmon, ⁵²W60, ⁵³Salmon-Salmon, ⁵⁴W60, ⁵⁵Salmon-Salmon, ⁵⁶W60, ⁵⁷Salmon-Salmon, ⁵⁸W60, ⁵⁹Salmon-Salmon, ⁶⁰W60, ⁶¹Salmon-Salmon, ⁶²W60, ⁶³Salmon-Salmon, ⁶⁴W60, ⁶⁵Salmon-Salmon, ⁶⁶W60, ⁶⁷Salmon-Salmon, ⁶⁸W60, ⁶⁹Salmon-Salmon, ⁷⁰W60, ⁷¹Salmon-Salmon, ⁷²W60, ⁷³Salmon-Salmon, ⁷⁴W60, ⁷⁵Salmon-Salmon, ⁷⁶W60, ⁷⁷Salmon-Salmon, ⁷⁸W60, ⁷⁹Salmon-Salmon, ⁸⁰W60, ⁸¹Salmon-Salmon, ⁸²W60, ⁸³Salmon-Salmon, ⁸⁴W60, ⁸⁵Salmon-Salmon, ⁸⁶W60, ⁸⁷Salmon-Salmon, ⁸⁸W60, ⁸⁹Salmon-Salmon, ⁹⁰W60, ⁹¹Salmon-Salmon, ⁹²W60, ⁹³Salmon-Salmon, ⁹⁴W60, ⁹⁵Salmon-Salmon, ⁹⁶W60, ⁹⁷Salmon-Salmon, ⁹⁸W60, ⁹⁹Salmon-Salmon, ¹⁰⁰W60, ¹⁰¹Salmon-Salmon, ¹⁰²W60, ¹⁰³Salmon-Salmon, ¹⁰⁴W60, ¹⁰⁵Salmon-Salmon, ¹⁰⁶W60, ¹⁰⁷Salmon-Salmon, ¹⁰⁸W60, ¹⁰⁹Salmon-Salmon, ¹¹⁰W60, ¹¹¹Salmon-Salmon, ¹¹²W60, ¹¹³Salmon-Salmon, ¹¹⁴W60, ¹¹⁵Salmon-Salmon, ¹¹⁶W60, ¹¹⁷Salmon-Salmon, ¹¹⁸W60, ¹¹⁹Salmon-Salmon, ¹²⁰W60, ¹²¹Salmon-Salmon, ¹²²W60, ¹²³Salmon-Salmon, ¹²⁴W60, ¹²⁵Salmon-Salmon, ¹²⁶W60, ¹²⁷Salmon-Salmon, ¹²⁸W60, ¹²⁹Salmon-Salmon, ¹³⁰W60, ¹³¹Salmon-Salmon, ¹³²W60, ¹³³Salmon-Salmon, ¹³⁴W60, ¹³⁵Salmon-Salmon, ¹³⁶W60, ¹³⁷Salmon-Salmon, ¹³⁸W60, ¹³⁹Salmon-Salmon, ¹⁴⁰W60, ¹⁴¹Salmon-Salmon, ¹⁴²W60, ¹⁴³Salmon-Salmon, ¹⁴⁴W60, ¹⁴⁵Salmon-Salmon, ¹⁴⁶W60, ¹⁴⁷Salmon-Salmon, ¹⁴⁸W60, ¹⁴⁹Salmon-Salmon, ¹⁵⁰W60, ¹⁵¹Salmon-Salmon, ¹⁵²W60, ¹⁵³Salmon-Salmon, ¹⁵⁴W60, ¹⁵⁵Salmon-Salmon, ¹⁵⁶W60, ¹⁵⁷Salmon-Salmon, ¹⁵⁸W60, ¹⁵⁹Salmon-Salmon, ¹⁶⁰W60, ¹⁶¹Salmon-Salmon, ¹⁶²W60, ¹⁶³Salmon-Salmon, ¹⁶⁴W60, ¹⁶⁵Salmon-Salmon, ¹⁶⁶W60, ¹⁶⁷Salmon-Salmon, ¹⁶⁸W60, ¹⁶⁹Salmon-Salmon, ¹⁷⁰W60, ¹⁷¹Salmon-Salmon, ¹⁷²W60, ¹⁷³Salmon-Salmon, ¹⁷⁴W60, ¹⁷⁵Salmon-Salmon, ¹⁷⁶W60, ¹⁷⁷Salmon-Salmon, ¹⁷⁸W60, ¹⁷⁹Salmon-Salmon, ¹⁸⁰W60, ¹⁸¹Salmon-Salmon, ¹⁸²W60, ¹⁸³Salmon-Salmon, ¹⁸⁴W60, ¹⁸⁵Salmon-Salmon, ¹⁸⁶W60, ¹⁸⁷Salmon-Salmon, ¹⁸⁸W60, ¹⁸⁹Salmon-Salmon, ¹⁹⁰W60, ¹⁹¹Salmon-Salmon, ¹⁹²W60, ¹⁹³Salmon-Salmon, ¹⁹⁴W60, ¹⁹⁵Salmon-Salmon, ¹⁹⁶W60, ¹⁹⁷Salmon-Salmon, ¹⁹⁸W60, ¹⁹⁹Salmon-Salmon, ²⁰⁰W60, ²⁰¹Salmon-Salmon, ²⁰²W60, ²⁰³Salmon-Salmon, ²⁰⁴W60, ²⁰⁵Salmon-Salmon, ²⁰⁶W60, ²⁰⁷Salmon-Salmon, ²⁰⁸W60, ²⁰⁹Salmon-Salmon, ²¹⁰W60, ²¹¹Salmon-Salmon, ²¹²W60, ²¹³Salmon-Salmon, ²¹⁴W60, ²¹⁵Salmon-Salmon, ²¹⁶W60, ²¹⁷Salmon-Salmon, ²¹⁸W60, ²¹⁹Salmon-Salmon, ²²⁰W60, ²²¹Salmon-Salmon, ²²²W60, ²²³Salmon-Salmon, ²²⁴W60, ²²⁵Salmon-Salmon, ²²⁶W60, ²²⁷Salmon-Salmon, ²²⁸W60, ²²⁹Salmon-Salmon, ²³⁰W60, ²³¹Salmon-Salmon, ²³²W60, ²³³Salmon-Salmon, ²³⁴W60, ²³⁵Salmon-Salmon, ²³⁶W60, ²³⁷Salmon-Salmon, ²³⁸W60, ²³⁹Salmon-Salmon, ²⁴⁰W60, ²⁴¹Salmon-Salmon, ²⁴²W60, ²⁴³Salmon-Salmon, ²⁴⁴W60, ²⁴⁵Salmon-Salmon, ²⁴⁶W60, ²⁴⁷Salmon-Salmon, ²⁴⁸W60, ²⁴⁹Salmon-Salmon, ²⁵⁰W60, ²⁵¹Salmon-Salmon, ²⁵²W60, ²⁵³Salmon-Salmon, ²⁵⁴W60, ²⁵⁵Salmon-Salmon, ²⁵⁶W60, ²⁵⁷Salmon-Salmon, ²⁵⁸W60, ²⁵⁹Salmon-Salmon, ²⁶⁰W60, ²⁶¹Salmon-Salmon, ²⁶²W60, ²⁶³Salmon-Salmon, ²⁶⁴W60, ²⁶⁵Salmon-Salmon, ²⁶⁶W60, ²⁶⁷Salmon-Salmon, ²⁶⁸W60, ²⁶⁹Salmon-Salmon, ²⁷⁰W60, ²⁷¹Salmon-Salmon, ²⁷²W60, ²⁷³Salmon-Salmon, ²⁷⁴W60, ²⁷⁵Salmon-Salmon, ²⁷⁶W60, ²⁷⁷Salmon-Salmon, ²⁷⁸W60, ²⁷⁹Salmon-Salmon, ²⁸⁰W60, ²⁸¹Salmon-Salmon, ²⁸²W60, ²⁸³Salmon-Salmon, ²⁸⁴W60, ²⁸⁵Salmon-Salmon, ²⁸⁶W60, ²⁸⁷Salmon-Salmon, ²⁸⁸W60, ²⁸⁹Salmon-Salmon, ²⁹⁰W60,

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*Lipid toxicity is T. battellae after 48 h, ⁵in vitro ER agonist potency determined using the solid rodentrogen screen (YES) and expressed as the equivalent response to 17 β -oestradiol (ng E2/E), *Malpighic activity determined using the Malpighic assay, ⁶in vitro AR agonist potency determined using the DR-CAUX assay and expressed as benz[a]pyrene (B[a]P) equivalent activity of the total extract over 24 h and 24 h, ⁷in vitro AR agonist potency determined using the DR-CAUX assay and expressed as 2,3,7,8-tetrachlorodibenzo[*a,h*]dioxin (TCDD) TEQ equivalent activity of the dioxin fraction over 24 h. Not analyzed.